



(19)

Europäisches Patentamt

European Patent Office

Office européen des brevets



(11)

EP 0 947 820 A2

(12)

EUROPEAN PATENT APPLICATION

(43) Date of publication:
06.10.1999 Bulletin 1999/40

(51) Int. Cl.⁶: G01N 1/38, G01N 35/00,
B01L 3/00

(21) Application number: 99103671.6

(22) Date of filing: 25.02.1999

(84) Designated Contracting States:
AT BE CH CY DE DK ES FI FR GB GR IE IT LI LU
MC NL PT SE
Designated Extension States:
AL LT LV MK RO SI

(30) Priority: 31.03.1998 EP 98105851

(71) Applicant:
Evotec BioSystems GmbH
22525 Hamburg (DE)

(72) Inventor: Henco, Karsten Dr.
D-40699 Erkrath (DE)

(74) Representative:
Meyers, Hans-Wilhelm, Dr.Dipl.-Chem. et al
Patentanwälte
von Kreisler-Setting-Werner
Postfach 10 22 41
50462 Köln (DE)

(54) Method for sample preparation

(57) The invention relates to a method for sample preparation comprising the steps of:

- combining a solution of a compound with an effective amount of an additive or a mixture of additives in a container,
- removing the solvent of said compound,
- storing said container,
- redissolving said compound,
- combining a solution of a substance to be tested for its potential interaction with said compound.

The invention further relates to a multi-well container, comprising:

- a plate defining wells, said wells having an open end, a substantially closed end and an inner part, said closed ends and said plate being formed of a material which is substantially impermeable to fluids, wherein
- the inner part of said wells being substantially coated with an effective amount of an additive or a mixture of additives, and
- wherein said additive or mixture of additives have taken up, or build an inclusion complex with, at least one type of compound.

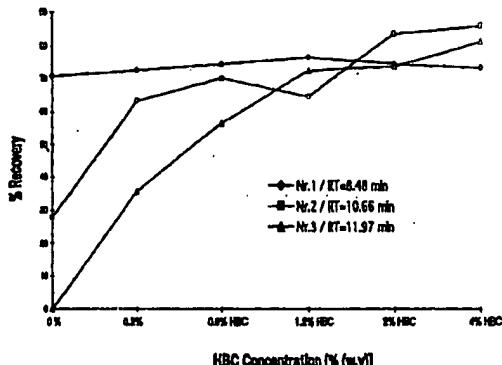


Figure 1

Description

[0001] The present invention relates to a method for sample preparation, to a multi-well container and to the use of additives in the production of multi-well containers.

5 [0002] Pharmaceutical companies nowadays apply high throughput drug discovery systems which enable automated, highly miniaturized screening of hundreds of thousands of samples to discover pharmacologically active compounds interacting with therapeutic targets on the molecular level. The compounds may be synthetic molecules - which are e.g. part of a compound library built by combinatorial chemistry or which are the collection of compounds synthesized manually during the companies history - or they may be isolated from natural sources such as bacteria or plants. There is

10 a strong need for a reliable early-on evaluation of the pharmacological potential of these compounds in order to determine the most promising candidates for the costly drug discovery. As a lot of compounds are available in only small amounts, it is of particular importance that these compounds can be stored in a safe manner without deterioration and further without a limited availability of the compounds during a screening assay e.g. due to their stickiness to the walls of a sample carrier.

15 [0003] It is state of the art to store the compounds to be screened in the form of concentrated stock solutions in solvents such as DMSO. Fractions of these stock solutions will then be transferred into sample containers in which a possible interaction between the compound and a target substance, e.g. a receptor of interest, will be tested in a so-called "screening assay". It cannot be excluded that DMSO might have unwanted or adverse effects on the assay. In addition, applicants' experimental results prove that some compounds are deteriorated or become instable during their storage

20 in DMSO.

[0004] Pharmaceutical formulations and food stuffs often contain additives such as cyclodextrins, different oligo- or polysaccharides or derivatives thereof for increasing their durability and bioavailability of their components.

25 [0005] Cyclodextrins are cyclic oligosaccharides. The most common cyclodextrins are α -cyclodextrin, which is composed of a ring of six glucose residues; β -cyclodextrin, which is composed of a ring of seven glucose residues; and γ -cyclodextrin, which is composed of a ring of eight glucose units. Cyclodextrins typically form a cavity which is essentially lipophilic, while the outside of the cyclodextrin is essentially hydrophilic; this combination of properties has led to widespread study of the natural cyclodextrins, particularly in connection with pharmaceuticals, and many inclusion complexes with drugs have been reported. Due to its cavity size, β -cyclodextrin has been of special interest, but its relatively low solubility in water (1.8 % w / v at 25 °C), attendant nephrotoxicity and hemolytic activity have limited its use in the pharmaceutical field.

30 [0006] Attempts to modify properties of the natural cyclodextrins have resulted in the development of a reasonably large number of cyclodextrin derivatives. In general, these chemically modified cyclodextrins are formed by reaction of the hydroxyl groups attached to the carbons 2, 3 or 6 without disturbing the α (1→4) hemiacetal linkages.

35 [0007] Inclusion complexes of α -, β - or γ -cyclodextrin or their mixtures with a variety of drugs have been described by numerous parties and various advantages have been attributed to the complexes. For instance, U.S. Patent 4,596,795 describes inclusion complexes of sex hormones, particular testosterone, progesterone and estradiol, with specific cyclodextrins, preferably hydroxypropyl- β -cyclodextrin and poly- β -cyclodextrin. Janssen Pharmaceutica N.V.'s International patent application No. PCT/EP 84/00417 (published under WO 85/02767) describes pharmaceutical compositions comprising inclusion compounds of drugs, which are unstable or only sparingly soluble in water, with partially

40 etherified β -cyclodextrin derivatives having hydroxyalkyl and optionally additional alkyl groups. Among the cyclodextrin derivatives contemplated is hydroxypropyl- β -cyclodextrin, while the drugs include nonsteroidal anti-rheumatic agents, steroids, cardiac glycosides and derivatives of benzodiazepine, benzimidazole, piperidine, piperazine, imidazole and triazole. The pharmaceutical compositions described include oral, parenteral and topical formulations, with 4 to 10 % solutions of cyclodextrin derivatives being used to solubilize the various drugs.

45 [0008] U.S. Patent 4,727,064 claims a composition containing an amorphous complex of cyclodextrin and a drug, and a method of producing a stabilizing amorphous complex of a drug and a mixture of cyclodextrins comprising (1) dissolving an intrinsically amorphous mixture of cyclodextrin derivatives which are water soluble and capable of forming Inclusion complexes with drugs in water; and (2) solubilizing lipophilic drugs into aqueous media to form a solution and form a solubilized drug/cyclodextrin complex. The patent describes the preparation of various substituted amorphous cyclodextrins, including hydroxypropyl- β -cyclodextrin and hydroxypropyl- γ -cyclodextrin. It is stated that the cyclodextrin additives may be utilized in weight percent of usually about 40 - 60 % of the drug solution composition and may be utilized from about 5 - 95 % of the drug solution composition.

50 [0009] U.S. Patent 5,580,856 discloses that dried proteins are stabilized against loss of biological activity in formulations by adding a reconstitution stabilizer upon rehydration of the dried protein.

55 [0010] Immobilized cyclodextrins may be obtained using a variety of procedures. U.S. Patent 5,183,809 provides a method of stabilizing protein factors, in particular heparin binding growth factor, by combining it with a polyionic derivative of a cyclodextrin polymer or a polyionic derivative of at least one cyclodextrin immobilized on a solid surface. These can be immobilized on a solid surface by covalently linking the derivatives e.g. via a linker arm. Polyanionic cyclodex-

trins can e.g. be impregnated on an inorganic xide which strongly binds anions. Polyionic derivatives may further be immobilized on a solid surface via electrostatic interactions between the ionic entities of the solid surface. The protein factor is bound to the cyclodextrin by contacting the protein factor suspended in a solution such as a low salt solution that promotes the binding of the protein factor to the cyclodextrin. The protein factor is released by contacting the protein factor/cyclodextrin polymer complex with a such or a solution such as a high salt solution that would promote the release of the protein factor.

[0011] A microtiterplate (MTP) for enzyme immunoassays is a known tray-like article having a plurality of wells which are usually aranged in a rectangular array, typically having 8 rows by 12 columns, and which are usually made by moulding from a transparent plastics material such as transparent polystyrene. In conventional use, e.g. ELISA, the tray including the wells is coated with an antigen or antibody either directly or indirectly and used to measure the degree of binding of the antibody with an enzyme conjugate by introducing the latter into the wells of the coated tray, incubating the two together in the tray, and sub-sequently measuring the optical density provided by the action of the enzyme on a substrate.

[0012] None of the foregoing references are believed to anticipate or render obvious applicants' invention as described and claimed herein.

[0013] The object of the present invention is to avoid the drawbacks of the prior art as adressed herein above. In contrast to the prior art - especially to the field of galenics where the object is to provide an additive which is suited to stabilize a well-defined drug - it is an object of the present invention to provide a solution for the storage of a high number of different compounds. These compounds often have different properties such as charge, hydrophobicity, molecular weight etc. The compounds may be synthetic molecules or they may be isolated from natural sources such as bacteria or plants. Especially for the purposes of high throughput screening, this high diversity of compounds with often unknown properties should be stored in a safe manner without the need of adjusting the additive to each compound.

[0014] The present invention relates to a method for sample preparation comprising the steps of:

- 25 - combining a solution of a compound with an effective amount of an additive or a mixture of additives in a container,
- removing the solvent of said compound,
- storing said container,
- redissolving said compound,
- combining a solution of a substance to be tested for its potential interaction with said compound. .

[0015] The present invention combines the following improvements: a) decreased tendency of the dry-stored compounds to stick to the walls of the sample carrier or to form insoluble aggregates during the drying process so that they can be resolved efficiently; b) dry-stored compounds can be easily shipped to another place where the actual screening process can be performed; c) low deterioration rate of the stored compounds; d) simpler plate handling; e) no humidity control, especially in minuaturized multi-well containers; f) the screening assay can be performed in aqueous solution.

[0016] Although the present invention may be used with compounds that are highly soluble in water, the fullest advantage of the solubilizing effect of additives will be realized in connection with compounds that have low solubility in aqueous solution.

[0017] The present invention has applications in high throughput screening (HTS) for pharmaceutically active compounds requiring efficient release/dissolution of intact ingredients in the presence of assay buffer. In addition, it is a valuable tool in diagnostics.

[0018] The method of the present invention requires removing the solvent of the compound. In one embodiment it further requires removing the solvent of an additive or a mixture of additives. Removing processes that can be used within the present invention have been described in the prior art and include, but are not limited to, drying processes such as lyophilization, spray-drying and air drying. A lyophilization cycle is usually composed of three steps: freezing, primary drying and secondary drying, as e.g. described in U.S. Patent 5,580,856. Spray-drying is typically achieved by microdispersing a solution into a stream of hot gas. Air-drying is typically performed by placing solutions at ambient temperatures in a very low humidity environment.

[0019] For the purpose of performing e.g. a screening assay, the method further comprises the step of identifying whether a compound interacts with said substance. The identification process is preferably based on the application of fluorescence techniques. It is preferred to use confocal fluorescence techniques such as fluorescence correlation spectroscopy (FCS), as in detail described in WO 94/16313 (herewith incorporated by reference) and WO 96/13744 (herewith incorporated by reference). It might further be preferred to rely on a fluorescence technique which is suited to determine molecular brightnesses and which is in detail disclosed in PCT/EP 97/05619 (herewith incorporated by reference).

[0020] In a further embodiment, the additive or a mixture of additives is present in the container before adding a solution of a compound. It might be particularly useful to use a container in which the additive or mixture of additives is present in immobilized form. Immobilization can e.g. be achieved by covalently coupling the additive to the container.

This embodiment might be of advantage as the risk of disturbance of the assay conditions due to the presence of additive is minimized. In a further embodiment, immobilization can also be due to non-covalent interaction between the additive and the walls of the container.

[0021] In some situations it might be preferred that a solution of an additive or mixture of additives is combined with said solution of a compound. For storage purposes, it is necessary that the solvent of said additive(s) is removed. This removal can preferably be performed together with the removal of the solvent of the compound.

[0022] In another embodiment, the step of redissolving the compound is carried out by adding a solution of a substance to be tested for its potential interaction with said compound.

[0023] For screening purposes, it is preferred to use a multi-well container such as a known microtiter or nanotiter plate. It is preferred to have wells with a volume smaller than 10 μ l, especially smaller than 2 μ l. Redissolution in these small compartment can be achieved without shaking or stirring.

[0024] Though a lot of different additives can be used with the present invention, it is particularly preferred to use a cyclodextrin or a derivative thereof. It is advantageous to use hydroxypropyl- β -cyclodextrin. This should preferably be present in a final amount being from about 0.05 % to 4 % by weight, based on the total volume of the final solution after the solution of the substance to be tested has been added. In a further embodiment, hydroxypropyl- β -cyclodextrin is present in a final amount being from about 0.05 % to 2 % by weight, based on the total volume of the final solution after the solution of the substance to be tested has been added. In another embodiment, hydroxypropyl- β -cyclodextrin is present in a final amount being from about 0.1 % to 1.5 % by weight, based on the total volume of the final solution after the solution of the substance to be tested has been added.

[0025] The present invention also relates to a multi-well container, comprising:

- a plate defining wells, said wells having an open end, an essentially closed end and an inner part, said essentially closed ends and said plate being formed of a material which is substantially impermeable to fluids, wherein
- the inner part of said wells being substantially coated with an effective amount of an additive or a mixture of additives, and
- wherein said additive or mixture of additives have taken up, or build an inclusion complex with, at least one type of compound.

[0026] It might be preferred to use multi-well containers disclosed in WO 93/07258 (herein incorporated by reference).
[0027] This patent application discloses a cell culture substrate having a microstructured, plate-like body with a plurality of depressions mutually separated by webs for receiving cells or cell aggregates.

[0028] The present invention further relates to the use of additives for the production of multi-well containers.

[0029] The nature and advantages of the present invention may be better understood on the basis of the following figures and examples.

Figure 1 shows the hydroxypropyl- β -cyclodextrin concentration dependence of recoveries after dry-storage.

Figure 2 shows the recovery of extract compounds.

Figure 3 shows the recovery of shelf-compounds.

Figure 4 shows the recovery of compounds in water and DMSO after dry-storage.

Figure 5 shows the degradation time-course of compound L.

Figure 6 shows the recovery of compounds after dry-storage in wells substantially coated with an effective amount of an additive and uncoated wells.

Figure 1 discloses the HBC concentration dependence of recoveries after dry-storage. 15 μ l aliquots of a natural extract were desiccated and stored in microtiter plates in the presence of increasing amounts of HBC (2-hydroxypropyl- β -cyclodextrin) as described in the legend to Fig. 2. The recovery of three representative peaks was determined. Retention-time of Peaks No. 1 to No. 3 were 8.48 min, 10.66 min and 11.97 min, respectively. The peaks represent a more hydrophilic, a less hydrophilic and a hydrophobic compound, respectively. "0% HBC" corresponds to the control, where water was added to the extracts instead of HBC solution. %-recovery values were determined as follows: Peak areas of the DMSO control were considered "100% recovery" (% Recovery = peak area of dried sample/peak area of DMSO control). The HPLC conditions were as follows:

Injection Volume: 10 μ l

Temperatur : Room-Temperature
 HPLC Säule: Merck LiCosphere RP-18,5 μ m
 Run-Time: approx. 11 min
 Data Recording: 200 nm - 250 nm (Diode Array Detector)

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Gradient Details:

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Time	0.1% TFA/Wasser	0.1% TFA/Acetonitrile	Flow
0 min	90%	10%	1 ml/min
5 min	0%	100%	1 ml/min
6 min	90%	10%	1 ml/min
8 min	90%	10%	1 ml/min

Figure 2 shows the recovery of extract components. The experimental procedure was as follows: 15 μ l aliquots of extracts (50% DMSO, 10 mM NaCl) contained in wells of microtiterplates (MTPs) were dried in a desiccator overnight at room-temperature. Two different regimens were used for the addition of additive (HBC): In the first procedure, HBC was coated to MTP-wells in advance by drying 1.5 μ l of a 15% (w / v) stock solution to the bottom of the well (CD coated). In the second procedure, the same volume of HBC solution was mixed with the extract solution before desiccation (CD mixed). After desiccation, pellets were resuspended in 15 μ l of water and centrifuged for 5 min at 15000 rpm. 10 μ l of the supernatant were HPLC-analyzed. 10 μ l of extract (DMSO solutions) served as control. HPLC conditions were identical to those described in the legend to Figure 1 (10 μ l injection volume).

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Figure 2A shows the HPLC profile of the extract. Recovery of 6 peaks (arrows) were compared under the conditions described above except gradient details. A linear acetonitrile/water gradient (0-100% acetonitrile in 15 min) was used.

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Figure 2B gives a graphic representation of recovery of the 6 peaks high-lighted in Fig. 2A in the presence or absence of 1.5% HBC.

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Figure 3 shows the recovery of shelf-compounds. Figure 3 a shows the HPLC profile of 10 μ l cocktail of 6 shelf-compounds (800 μ M each). Retention-times: 1: 7.3 min; 2: 8.5 min; 3: 10 min, 4: 11 min; 5: 12 min; 6: 13.3 min. Figure 3b discloses a graphic representation of recovery observed in the presence or absence of 1.5% HBC for each compound. The experimental procedure was as described for the extract components (Fig. 2).

40

Figure 4 shows the recovery of compounds in water and DMSO after dry-storage. 20 μ l aliquots of a panel of 20 reference compounds (1 mM each in 50 % DMSO, 10 mM NaCl) were stored in microtiter plates in the presence and absence of additives (HBC, HGC, Trehalose; 1.5% (w / v) each) for 10 weeks. The dried compounds were reconstituted by the addition of 20 μ l water (or 100% DMSO). In order to actively support the dissolution process, the 20 μ l were pipetted up-and-down several times. Insoluble particles were sedimented by centrifugation. Recoveries were determined by comparing the peak areas of the stored compound with those of the -80 °C controls (the retention times of each compound is also shown).

45

Figure 5 shows the degradation time-course of compound L. HPLC profiles of compound L stored under different conditions are shown: Control profiles (-80 °C storage), storage as DMSO solutions (4 weeks, 10 weeks) and desiccated storage for 10 weeks (without additive and in presence of 1.5% (w / v) HBG. Compound L, as deduced from the control stored at -80 °C, elutes as one main peak at 4.5 min. The top profiles show the results obtained for storage at 4 °C. Fig. 4B shows the results for room-temperature storage. The shaded areas highlight the position of the peak of the intact compound (-80 °C control used as reference). The shaded area at approximately 2 min. highlights the region of a broad peak, probably a degradation product of compound L.

50

Figure 6 shows the recovery of compounds after dry-storage in wells substantially coated with an effective amount of an additive and uncoated wells. Figure 6 a shows the RP-HPLC profile of the separation of a mixture of tetramethylrhodamine glycine and its reaction products using a linear gradient of water (plus 0.1%TFA) and acetonitrile from 0% to 100% acetonitrile in 15 minutes, monitoring the absorption at 540 nm. Figure 6b and 6c disclose the recovery of this mixture after dry-storage and subsequent dissolution in microtiter plates (6b) precoated with 2-hydroxypropyl- β -cyclodextrine and (6c) uncoated plates. The measurements were done by fluorescence correlation spectroscopy at 543 nm.

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Example 1: Long term stability studies of 20 compounds

[0029] The aim of these experiments was to monitor the stability of a panel of 20 reference compounds stored under different conditions over a period of months. Some of the compounds were expected to be instable when stored as DMSO solutions. Dry-storage of compounds in the presence and absence of additives was therefore of particular interest.

Treatment & Storage

[0030] Compounds were received as solids. Details of the compounds are listed in Table 1. For convenience, compounds were alphabetically labelled A-T. 10 mM stock solutions were prepared by dissolving them in appropriate amounts of 100 % DMSO and stored until further use, i.e. until the actual storage experiments, at -80°C.

Table 1

Compounds		
Code	Retention time / min.	Molecular weight
A	5.55	420.24
B	3.55	232.29
C	5.14	292.38
D	4.96	364.93
E	4.21	312.39
F	2.61	175.24
G	3.31	361.85
H	4.53	468.28
I	2.3	387.85
J	5.07	488.61
K	3.86	324.43
L	4.44	420.54
M	4.67	448.40
N	5.27	414.03
O	1.01	267.25
P	3.8	295.80
Q	4.27	422.40
R	4.33	307.80
S	4.52	370.45
T	2.62	255.80

[0031] Prior to storage experiments, HPLC profiles of the 10 mM stock solutions were recorded. 20 nmol of each compound was found to be sufficient to yield interpretable HPLC profiles. Thus, aliquots of at least 20 nmol were stored.

[0032] Storage conditions differed in:

1. Storage Temperature: 4 °C and room-temperature "RT" (Control: -80 °C storage)
2. Additives: presence and absence of additives (hydroxypropyl- β -cyclodextrin, abbr. "HBC"; hydroxypropyl- γ -cyclodextrin, abbr. "HGC"; Trehalose)
3. „Dry-storage" and storage as „DMSO-solution" (50 % DMSO, 10 mM NaCl)

[0033] A complete list of the storage conditions for each compound is shown in Table 2 below. Generally, the com-

pounds were stored in the dark, however, one aliquot of compound/DMSO solution was stored at day-light (Condition 2).

5

Table 2

Storage Conditions			
Condition	Temp.	Desiccated / Solution	Additive
10	1	-80 °C	"Control"
	2	RT	Solution (daylight)
	3	RT	Solution
	4	RT	Solution
15	5	RT	Desiccated
	6	RT	Desiccated
	7	RT	Desiccated
	8	RT	Desiccated
20	9	4 °C	Solution
	10	4 °C	Solution
	11	4 °C	Desiccated
25	12	4 °C	Desiccated
	13	4 °C	Desiccated
	14	4 °C	Desiccated

30 [0034] Commercial sources of additives and buffer components are listed in Table 3.

Table 3

Commercial Sources of Additives and Fine Chemicals				
Name	Supplier	Cat. No.	Lot Number	
Sodium Chloride (NaCl)	Merck	1.06404	K22156904533	
Hydroxypropyl- β -cyclodextrin (HBC), 45 % (w/v) solution	RBI	H-107	BJO-996A	
Hydroxypropyl- γ -cyclodextrin (HGC), 45 % (w/v) solution	RBI	H-125	CW-190A	
Trehalose	Sigma	T-5251	16H3816	
Dimethylsulphoxide (DMSO)	Merck	1.02952	K22605452609	

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Storage Procedures and Containers

1. "-80 °C Controls"

50 [0035] Of each compound several 20 μ l aliquots were stored at -80 °C in Eppendorf tubes. At the time of analysis one aliquot was removed, thawed and transferred to a NPLC vial. Although the -80 °C controls were always the first samples to be HPLC analyzed, they resided up to 2 hours at room-temperature in the HPLC-autosampler.

55

2. "dry-storage"

[0036] 20 μ l aliquots of each compound (1 mM, contained in 50 % DMSO, 10 mM NaCl) were placed in the wells of Greiner V-shaped microtiterplates (MTPs, Greiner Product Code: 651101) and dried down in a desiccator (overnight at room-temperature). The MTPs containing the dried pellets were covered with a transparent, self-adhesive film (Greiner,

5 CatNo: 676001) and were placed in made-to-measure heat-sealable multi-layer storage bags (outside layer: aluminium; Inside layer: polyethylene). To ensure a dry atmosphere inside the bag a small amount of blue silica-gel was added prior to sealing. For HPLC analysis, the MTPs were removed from the bags and the compounds reconstituted in 20 µl of water by pipetting up-and-down several times. In order to sediment any undissolved particles, the MTPs were centrifuged at 3700 x g 5 min and the supernatants were transferred to HPLC vials.

3. "DMSO-solutions"

10 [0037] DMSO-solutions of compounds (1 mM, contained in 50 % DMSO, 10 mM NaCl) were also stored in MTPs (160 µl/well) which were sealed in storage bags (without silica gel). For storage at day-light, the MTP was covered with transparent film and placed on top of the laboratory shelf (NB: Recently, transmission spectra of the sealing foil were recorded. They reveal a sharp cut-off below 300 nm). For HPLC analysis, a 20 µl aliquot of each compound was removed and the MTP was subsequently resealed in a new bag.

15 Recipes of Solutions containing Compounds & Additive

20 [0038] In order to reduce the number of pipetting steps a set of additive stock solutions were prepared (Table 4) and used to create „Master-mixes“ containing compounds. The volumes shown in Table 4 and of the „Master-mixes“ are calculated for one panel of 10 compounds (e.g. compounds A - J).

Table 4

Stock Solutions of Additives and NaCl Solutions	
Stock Solution	Composition
20 mM NaCl (7ml)	140 µl 1M NaCl + 6860 µl water
40 mM NaCl (6ml)	240 µl 1M NaCl + 5760 µl water
6% HBC (2.3ml)	307 µl 45% (w/v) HBC Solution + 1993 µl water
6% HGC (1.5ml)	89.7 mg HGC + 1495 µl water
6% Trehalose (1.9 ml)	111.1 mg Trehalose + 1852 µl water

35 Master-Mix 1: Storage without Additive

[0039] Final Composition: 1 mM compound, 50% (v / v) DMSO / 10mM NaCl

40 1. Premix: 5111 µl 100% DMSO + 6389 µl 20 mM NaCl
 2. Aliquot into 10 Eppendorf tubes (1080 µl each).
 3. For each of the 10 compounds, 120 µl stock solution (10 mM in 100% DMSO) were added to yield a total of 1.2 ml of 1 mM compound solution.

[0040] 1.2 ml solution were used for 7 batches:

45 > 21 x 20 µl aliquots: 7 batches x 3 different conditions: -80 °C, dry-storage at 4 °C & RT
 > 3 x 160 µl aliquots for storage in DMSO-solution at RT, RT-light, 4°C

Master-Mix II: Storage in presence of 1.5% (w/v)HBC

50 [0041] Final composition: 1 mM compound, 50% (v / v) DMSO / 10mM NaCl / 1,5% (w / v) HBC

1. Premix: 3556 µl 100% DMSO + 2222 µl 40mM NaCl + 2222 µl 6% (w/v) HBC
 2. Aliquot (720 µl each) in 10 Eppendorf tubes
 3. For each of the 10 compounds, 80 µl stock-solution (10 mM in 100 % DMSO) were added

[0042] 0.8 ml stock solution were used for 7 batches:

> 14 x 20 µl aliquots (two different storage conditions): dry-storage 4°C & RT
 > 2 x 160 µl aliquots: storage in DMSO-Solution (RT, 4°C)

Master-Mix III: Storage in presence of 1.5% (w/v) HGC

5 [0043] Final composition: 1 mM compound, 50% (v/v) DMSO / 10mM NaCl /1.5% (w/v) HGC

1. Premix: 2222 µl 100% DMSO + 1389 µl 40mM NaCl + 1389 µl 6% HGC
2. Aliquot (360 µl each) in 10 Eppendorf tubes
- 10 3. For each of the 10 compounds, 40 µl stock-solution (10 mM in 100% DMSO) were added.

[0044] 0.4 ml stock solution were used for 7 batches:

> 14 x 20 µl aliquots (two different conditions): dry-storage 4 °C & RT

15 Master-Mix IV: Storage in presence of 1.5% (w/v) Trehalose

[0045] Final composition : 1 mM compound, 50% (v/v) DMSO / 10mM NaCl /1.5% (w/v) Trehalose

- 20 1. Premix 2222 µl 100% DMSO + 1389 µl 40mM NaCl + 1389 µl 6% Trehalose
2. Aliquot (360 µl each) in 10 Eppendorf tubes
3. For each of the 10 compounds, 40 µl stock-solution (10 mM in 100 % DMSO) were added.

[0046] 0.4 ml stock solution were used for 7 batches:

25 >14 x 20 µl aliquots (two different conditions): dry-storage 4 °C & RT

Example 2: Stability study of 3'Azido-3'deoxythymidine (AZT)

30 [0047] The method, according to claim 1 was used for a primer extension assay using HIV Reverse Transcriptase (HIV-RT). Precoating the assay plate: 1 µl aliquots of 0.1% (w/v) aqueous solution of HBC (2-hydroxypropyl-β-cyclodextrin) was pipetted into the wells of a nanotiter plate and subjected to a flow of nitrogen gas until all the solvent (water) was evaporated. The evaporation took place at room-temperature and resulted in the formation of a film consisting of amorphous HBC residing at the bottom of the nanotiter plate. As a control, a section of the nanotiter plate was not coated with HBC. The plate was subsequently loaded with 1 µl (1 mg/ml) of the HIV Reverse Transcriptase Inhibitor AZT (3'Azido-3'deoxythymidine) contained in 50 mM DMSO/ 10 mM NaCl. The solvent (DMSO/water) evaporated by exposure to a flow of nitrogen gas at room-temperature. As a control, some wells of the nanotiter plate (coated and non-coated) were not charged with AZT. The plates were stored in a desiccator overnight at room-temperature.

35 [0048] The primer extension assay was performed by pipetting 900 nanoliter of a reaction mix containing: 2 pmol TMR-labelled 20 base oligo deoxynucleotide annealed to a 200 base RNA-template (gag region of the HIV genome), 50 mM Tris-HCl pH 8.2, dissolved, 50 mM NaCl, 1 mM of each dNTP. The nanotiter plate was incubated at 40°C for 2 min and the extension reaction stopped by the addition of 100 nanoliter 0.5 M EDTA. The primer extension products were analyzed by FCS.

40 45 Results:

[0049]

50 1. A good primer extension (due to HIV-RT activity) was observed by FCS by the appearance of labeled DNA-strands of slow diffusion times, both in wells without AZT which were coated with HBC and in those not precoated with HBC.

55 2. Primer extension was found to be inhibited in wells which were coated with both HBC and AZT but only partially inhibited in wells not coated with HBC. The presence of HBC during dry-storage therefore preserves the inhibitory activity of AZT compared to dry-storage of AZT without any auxiliary substance.

[0050] The invention described and claimed herein is not to be limited in scope by the specific embodiments herein

disclosed, since these embodiments are intended as illustrations of several aspects of the invention. Any equivalent embodiments are intended to be within the scope of this invention. Indeed, various modifications of the invention in addition to those shown and described herein will become apparent to those skilled in the art from the foregoing description. Such modifications are also intended to fall within the scope of the appended claims.

5

Claims

1. A method for sample preparation comprising the steps of:

- 10 - combining a solution of a compound with an effective amount of an additive or a mixture of additives in a container,
- removing the solvent of said compound,
- storing said container,
- redissolving said compound,
- 15 - combining a solution of a substance to be tested for its potential interaction with said compound.

2. The method of claim 1, further comprising the step of identifying whether said compound interacts with said substance.

20 3. The method of claim 1, wherein said additive or said mixture of additives is present in said container before adding said solution of a compound, preferably in immobilized form.

25 4. The method of claim 1, wherein a solution of an additive or mixture of additives is combined with said solution of a compound and the solvent of said additive(s) is removed, preferably together with the solvent of said compound, before storing said container.

5. The method of claim 1, wherein the step of redissolving said compound is carried out by combining a solution of a substance to be tested for its potential interaction with said compound.

30 6. The method of claim 1, wherein said container is a multi-well container.

7. The method of claim 1, wherein said additive is cyclodextrin or a derivative thereof, such as hydroxypropyl- β -cyclodextrin.

35 8. The method of claim 7, wherein said hydroxypropyl- β -cyclodextrin is present in a final amount being from about 0.05 % to 4 % by weight, preferably from about 0.05 % to 2 % by weight, more preferably from about 0.1 % to 1.5 % by weight, based on the total volume of the final solution after the solution of the substance to be tested has been added.

40 9. A multi-well container, comprising:

- 45 - a plate defining wells, said wells having an open end, a substantially closed end and an inner part, said substantially closed ends and said plate being formed of a material which is impermeable to fluids, wherein
- the inner part of said wells being substantially coated with an effective amount of an additive or a mixture of additives, and
- wherein said additive or mixture of additives have taken up, or build an inclusion complex with, at least one type of compound.

50 10. The container of claim 9, wherein said additive is cyclodextrin or a derivative thereof, such as hydroxypropyl- β -cyclodextrin.

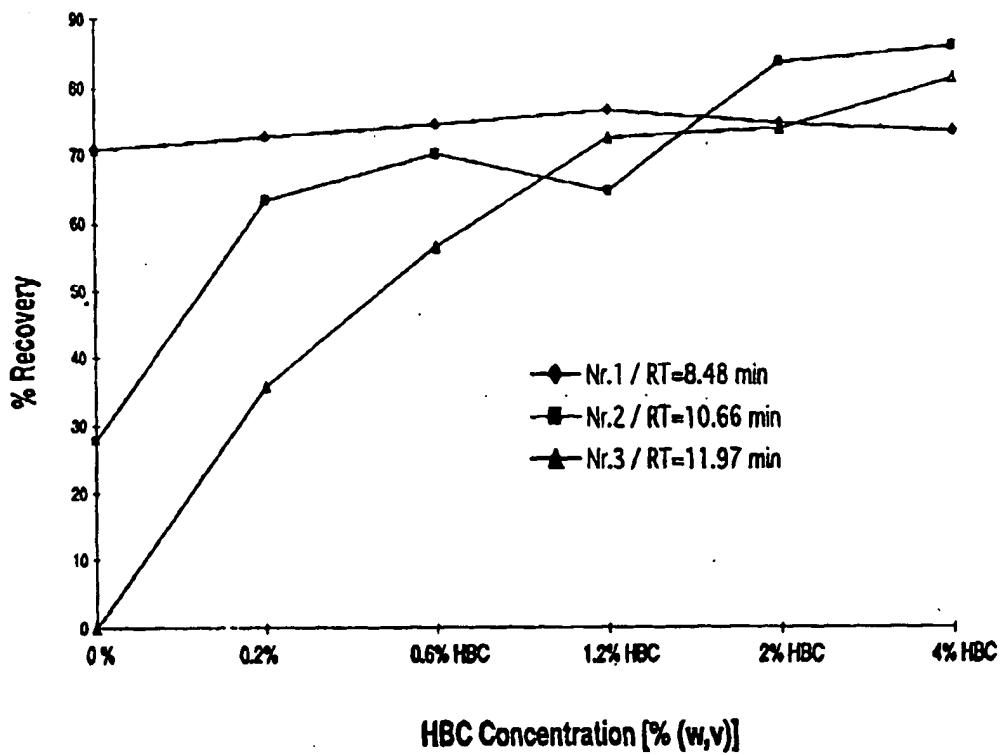


Figure 1

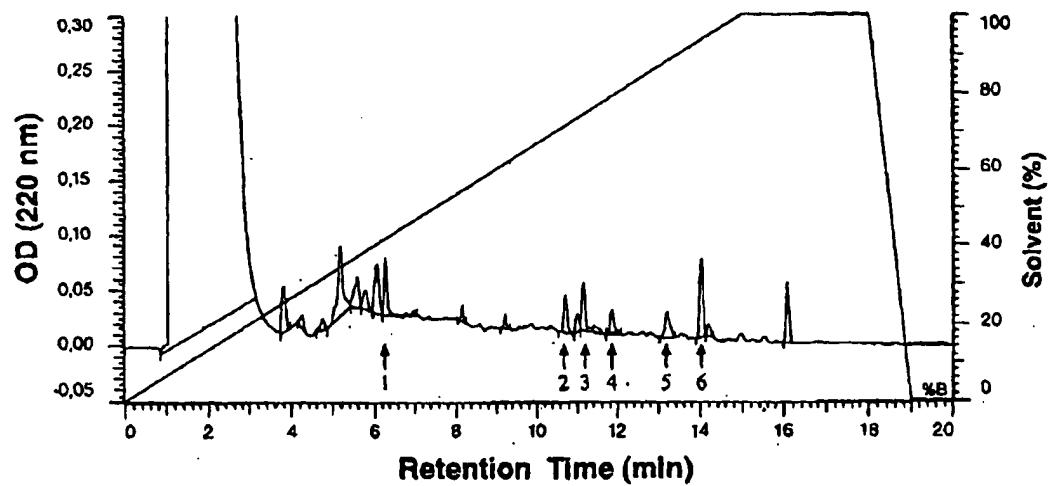
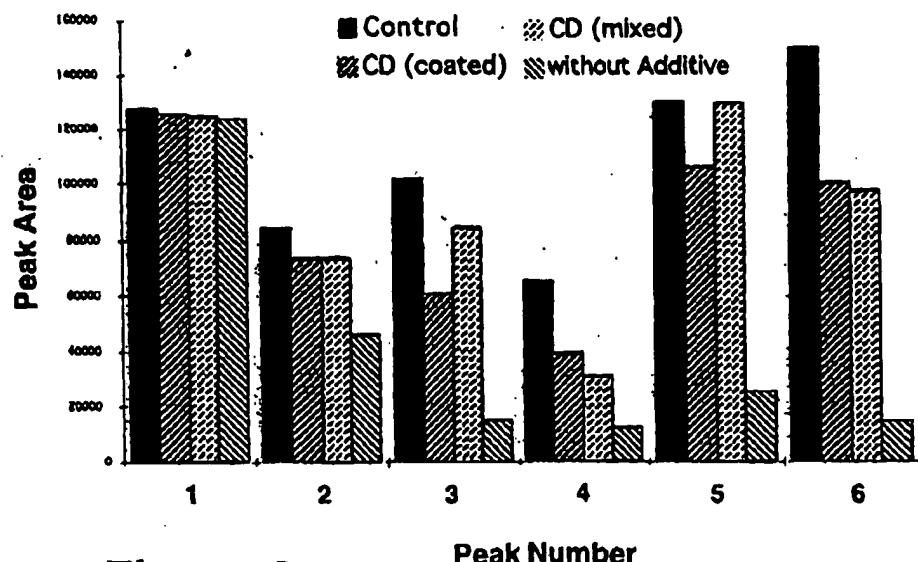
Fig. 2a**Fig. 2b****Figure 2**

Fig. 3a

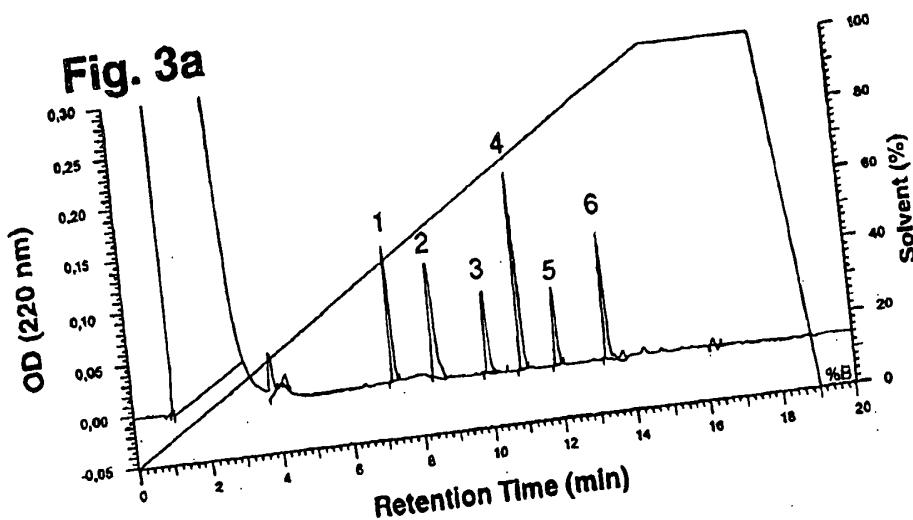


Fig. 3b

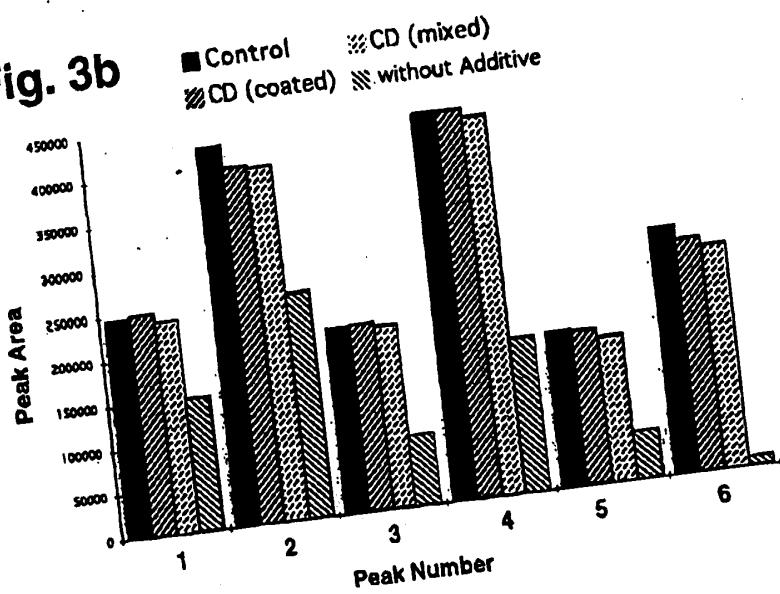


Figure 3

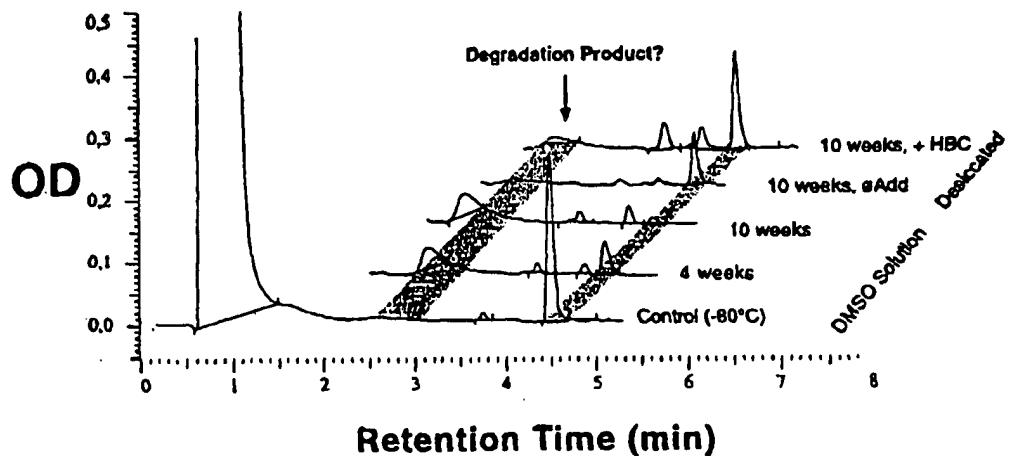
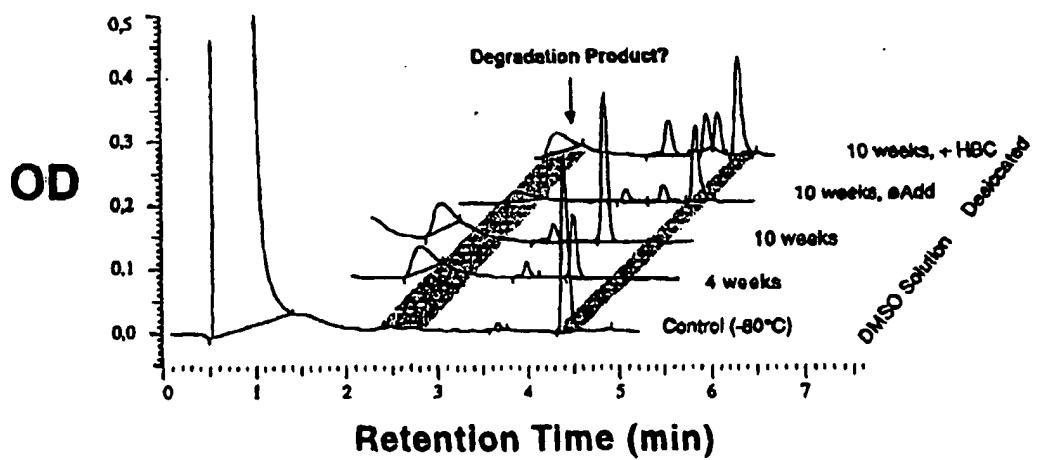
EVOTEC	Ret.	Water 4°C	Water RT	Water 4°C/RT	DMSO 4°C	DMSO RT
Code	(min)	ø Add	ø Add	HBC/HGC/Tre	ø Add	ø Add
A	5.55	***	**	****	****	****
B	3.55	****	****	****	****	****
C	5.14	***	***	****	****	****
D	4.96	****	****	****	****	****
E	4.21	*	****	****	**	****
F	2.61	****	****	****	****	****
G	3.31	****	****	****	****	****
H	4.53	**	**	****	***	***
I	2.3	****	****	****	****	****
J	5.07	*	*	HBC/HGC	*	*
K	3.86	**	**	****	***	***
L	4.44	Lower due to degradation				
M	4.67	*	*	HBC/HGC	****	****
N	5.27	****	****	****	****	****
O	1.01	****	****	****	****	****
P	3.8	****	****	****	****	****
Q	4.27	****	****	****	****	****
R	4.33	****	****	****	****	****
S	4.52	*	**	****	*	*
T	2.62	****	****	****	****	****

Recoveries

- **** >80%
- *** 60%-40%
- ** 40%-20%
- * 20%-0%

(100% = -80°C Control)

Figure 4

Fig. 5a**Fig. 5b****Figure 5**

